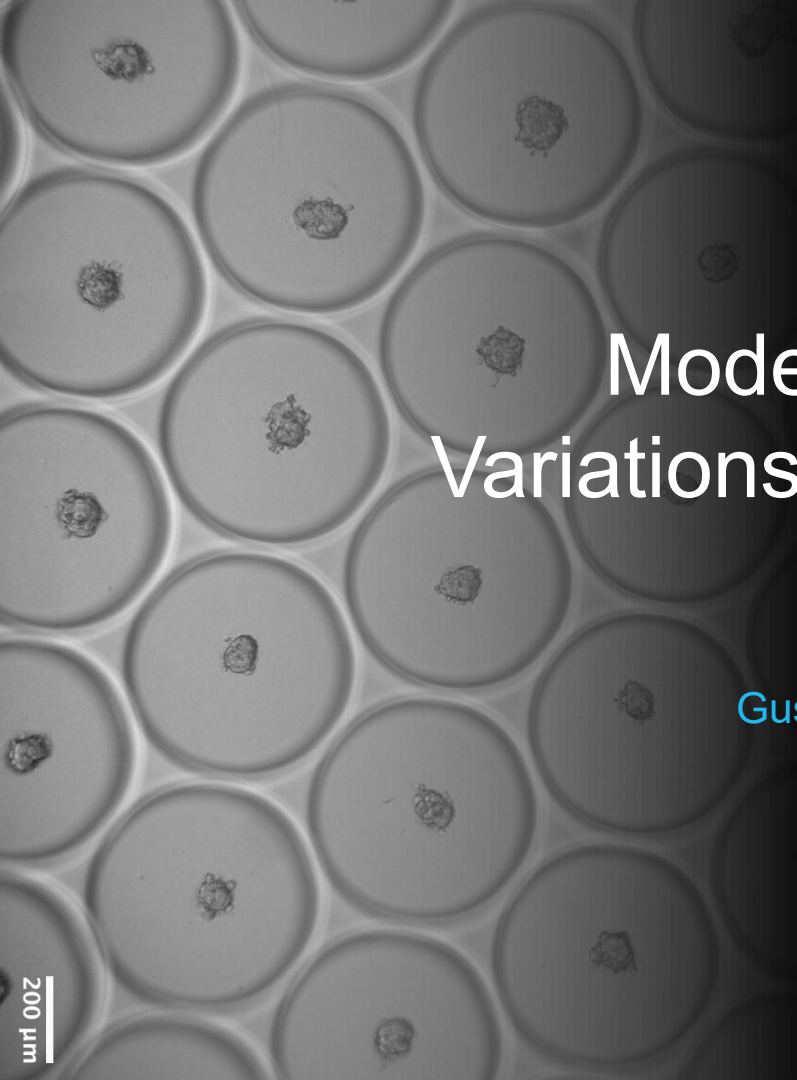


A grayscale micrograph showing a grid of circular wells in a multi-well plate. Each well contains a small, dark, irregularly shaped organoid. The organoids vary slightly in size and texture. The background is a light gray, and the wells are separated by thin, darker lines.

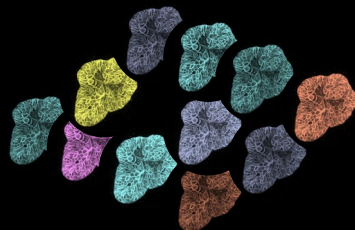
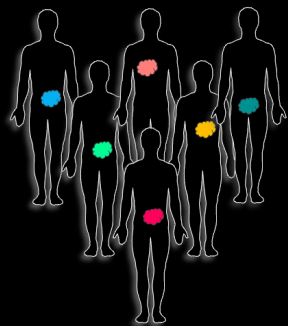
Modelling cancers, Variations around organoids

Fanny Jaulin
Gustave Roussy Institute

September 14th, 2023

Inserm





Cancer is a heterogenous
& complex tissue disease

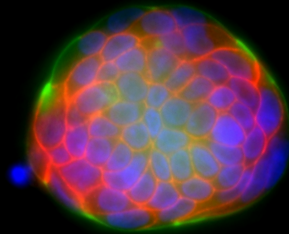
Relevant cancer models

To generate new knowledge

To improve translatability to patients

Underperforming oncology R&D
4% of drug candidates validated in the clinic

1- Modelling cancers

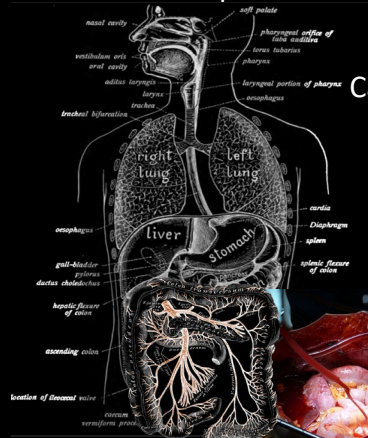


Translational Cell Biology

prospective systematic analyses
to reveal cancer cell behaviours

Bedside-to-bench

Cancer patients



Live Primary
Cancer specimens



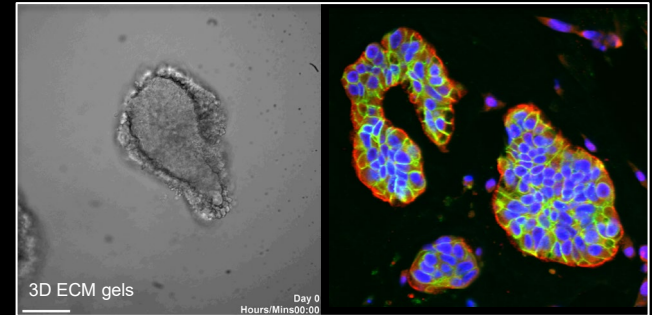
Surgery



Digestive Cancers

Digestive Pathologies committee

Explants for immediate and short-term studies



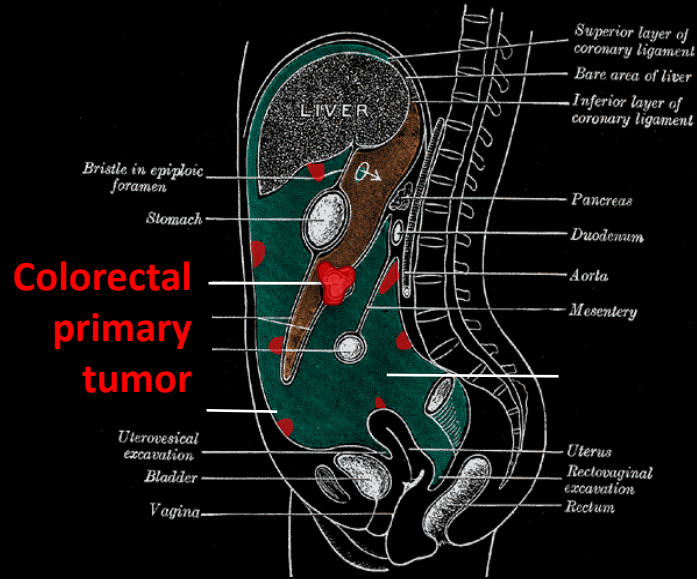
=> New and relevant knowledge

Bench-to-bedside

Biomarkers, Therapeutic targets

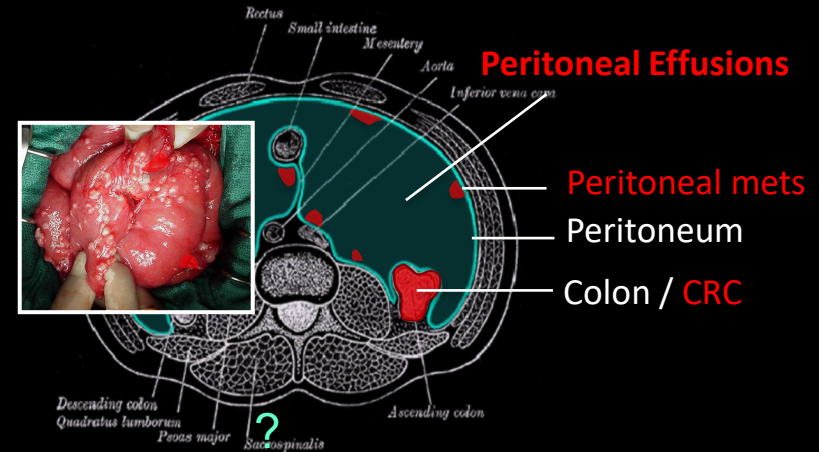
Hollistic approaches from large cohorts of CRC patients

>20 CRC patients at early stage



Libanje et al. EMBO 2019

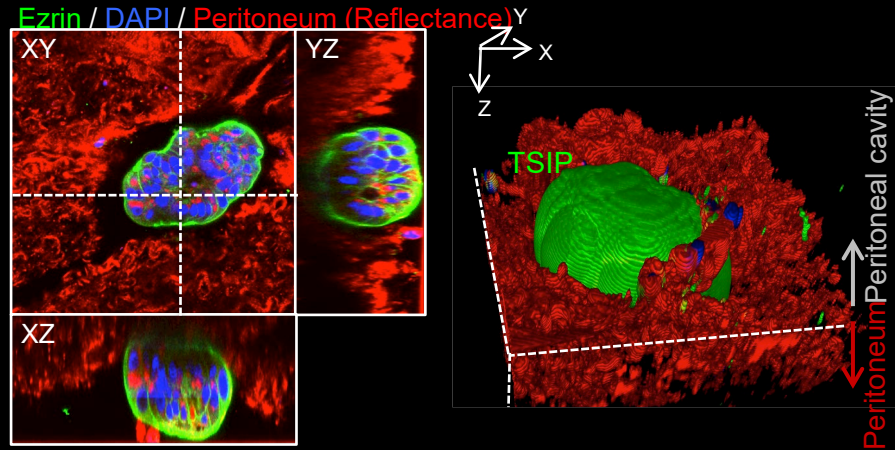
>50 CRC patients with advanced metastatic disease



Zajac et al. Nat Cell Biol 2018

Hollistic approaches from large cohorts of CRC patients

- Pros: Account for complexity & heterogeneity
Generate new & relevant knowledge (TSIPs, collective amoeboid migration)
- Cons: Complicated mechanistic studies
Does not “capitalize” on this precious resources to build collection



« All Models are wrong,
but some are useful »

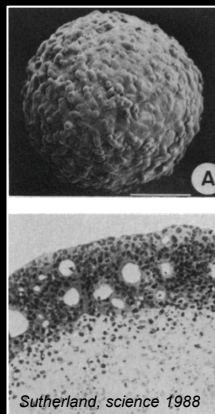


George Box
(1919 – 2013)

Ex vivo model systems for Research

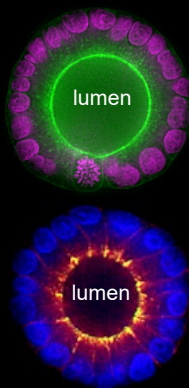
3D Models

Spheroids



80's

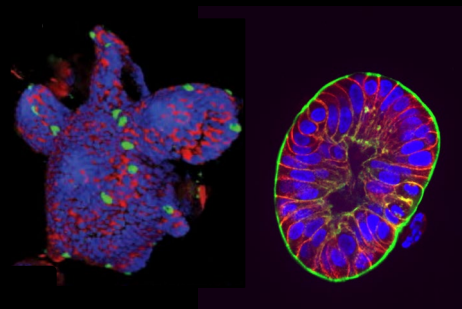
Cysts/Acini



90's

(M. Bissell, J Brugge)

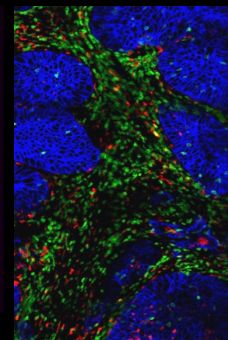
Organoïds /Tumoroids



2009

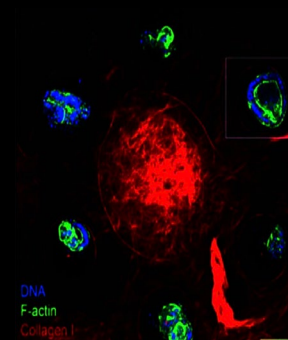
(Clevers, Knoblich)

Tissue slices



(E Donnadieu)

Organ-on-Chip



(S Decroix)



Vivo

2D/stiff culture

Artificial substrates

Physiological hydrogels (ECM-based)

Native tissue

Microfluidics

Culture system

Cell lines

Stem cells / Tumor fragments

Multiple cell types

Cells

—

+

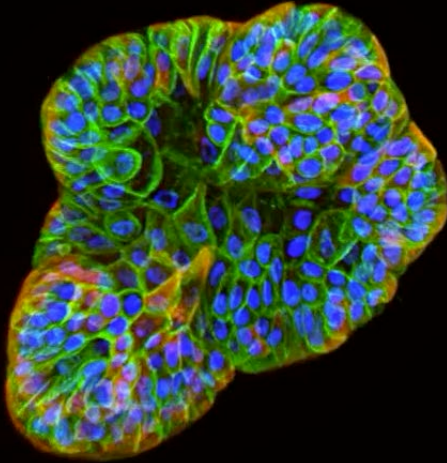
+

++

+/++

TME

Patient-derived (tumor) organoids -PDOs-



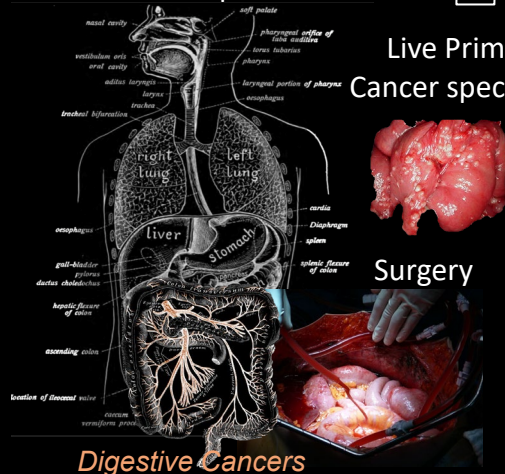
- Live specimen amplified *ex vivo* “indefinitely”
- Cryopreserved to build large collections capturing cancer heterogeneity
- Most faithful tumor avatar to date
 - Molecular make-up
 - Behaviors (vs explants)
- Versatile & scalable at the population level

Translational Cell Biology

prospective systematic analyses
to reveal cancer cell behaviours

Bedside-to-bench

Cancer patients



Live Primary
Cancer specimens



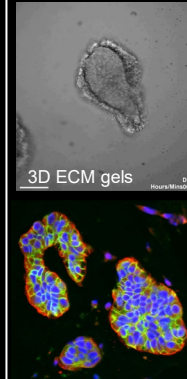
Surgery

Digestive Pathologies committee

Bench-to-bedside

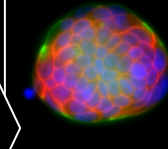
Biomarkers, Therapeutic targets

Explants



Generation

Biological collections (>120 PDOs)



Banked



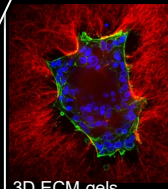
WES/RNAseq



Histology



Experiments!

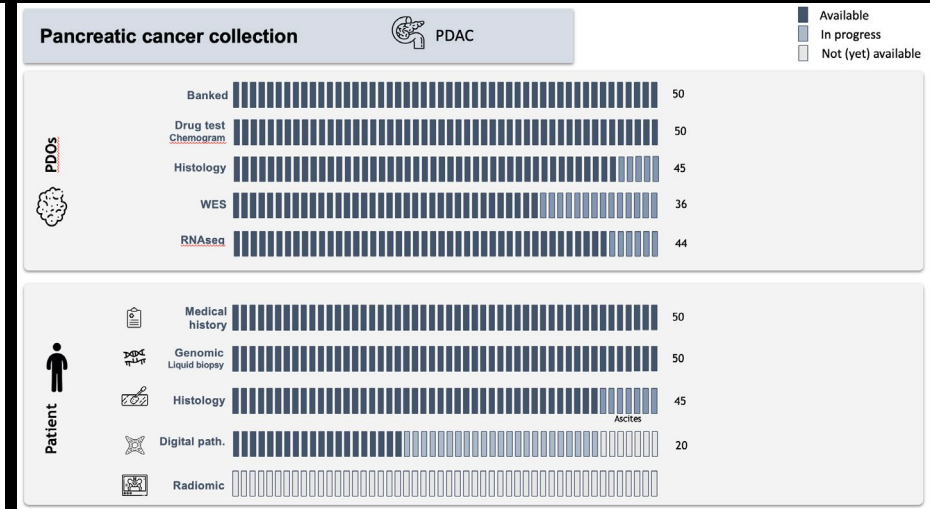
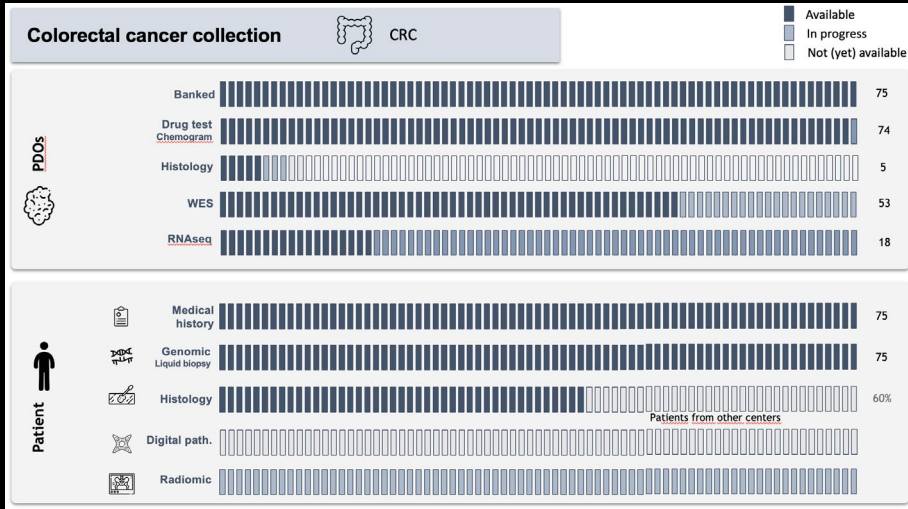


3D ECM gels

Validation

Hypotheses

Ex vivo tumor avatars collection that captures cancer complexity & heterogeneity at the population scale

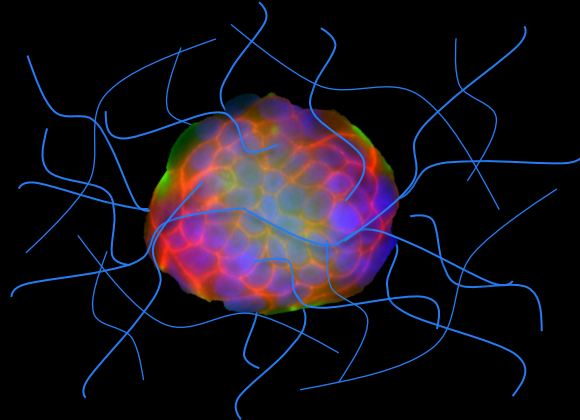


2- Fundamental research: tumor cell invasion

Collagen-I matrices

Extra-cellular matrix represents
up to 60% of the tumor mass

Henke et al, 2020



Simple collagen-I gel reveals invasive behaviors from explants and PDOs



Explants
& PDOs

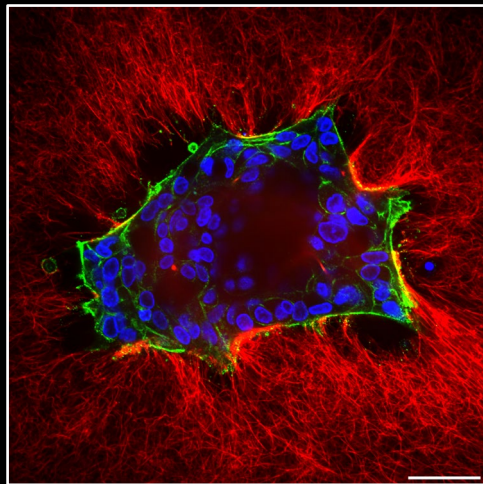


Collagen-I
hydrogel



Monitor
invasion

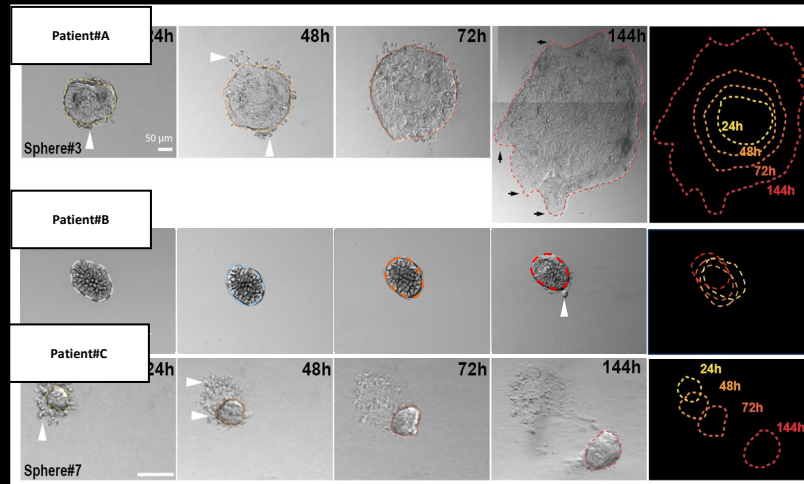
=> Profiling patterns reveals invasive behaviors
& tumor heterogeneity



PDO

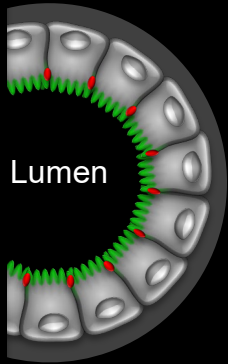


Primary explant

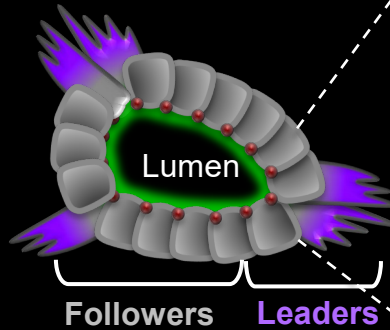


ROCK2 inhibition triggers leader cell formation & collective invasion

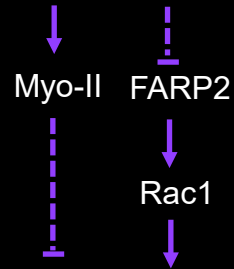
CRC "In situ"



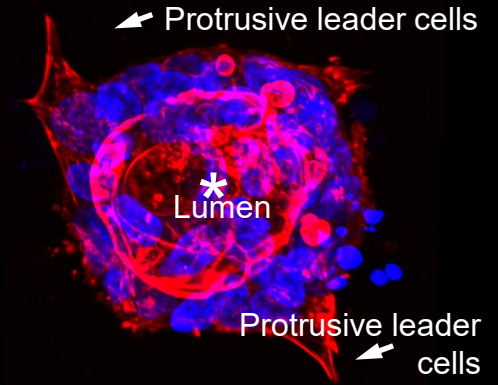
Collective Invasion



ROCK2

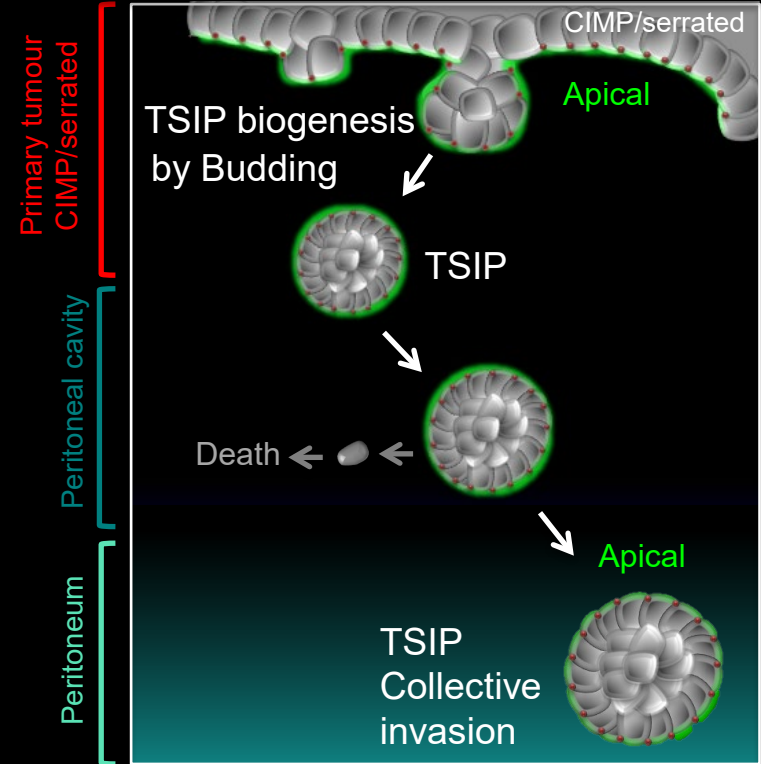
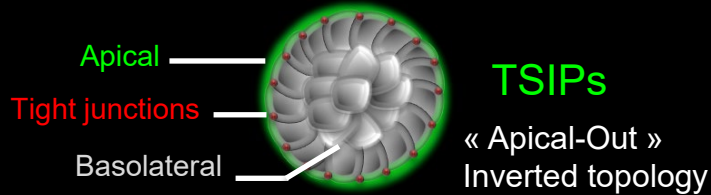
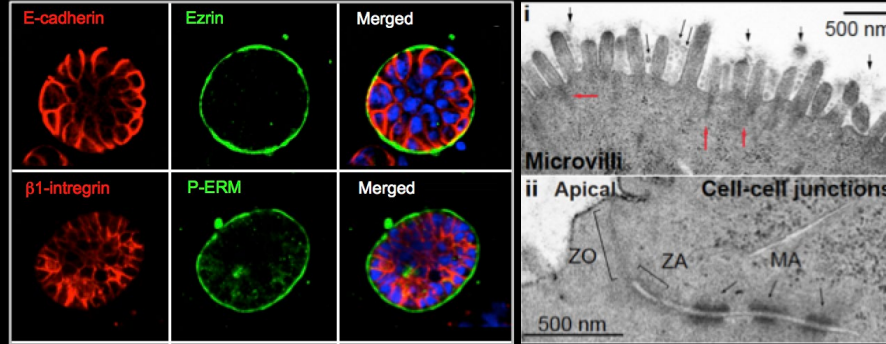


Leader cell formation

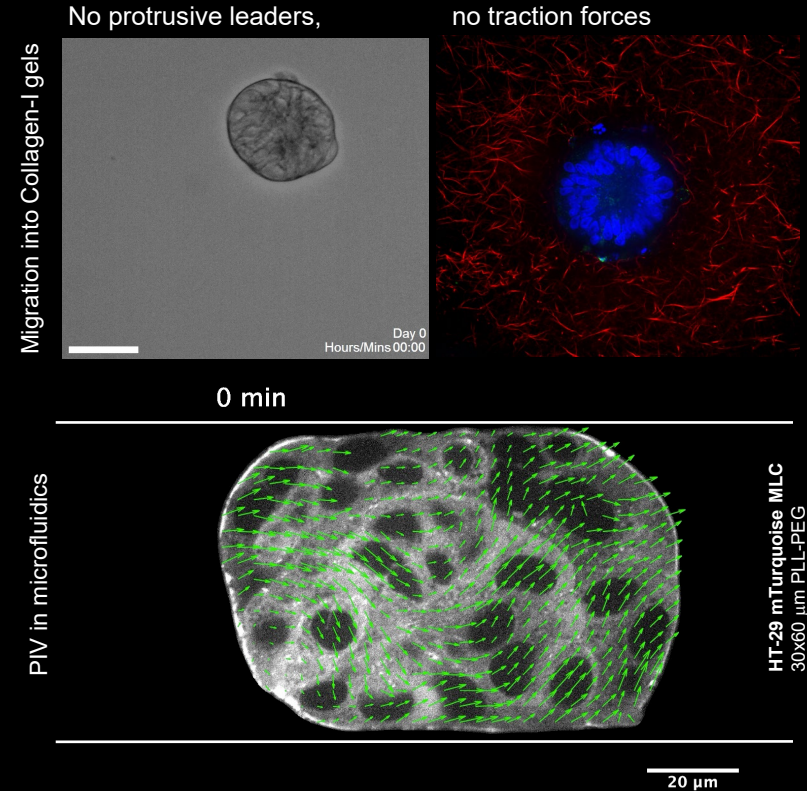
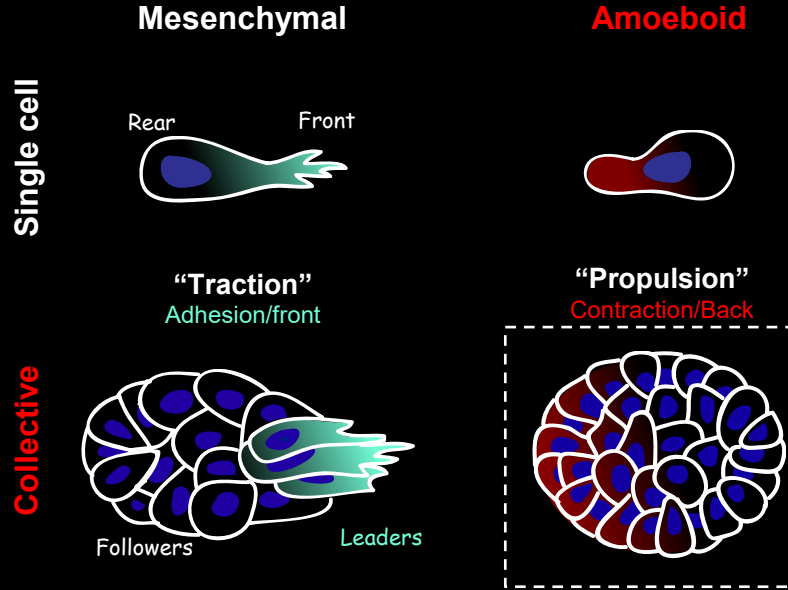


TSIPs are malignant tumor intermediates underlying the metastatic spread of serrated hypermethylated CRCs

50% of effusions from metastatic CRC patients contain TSIPs



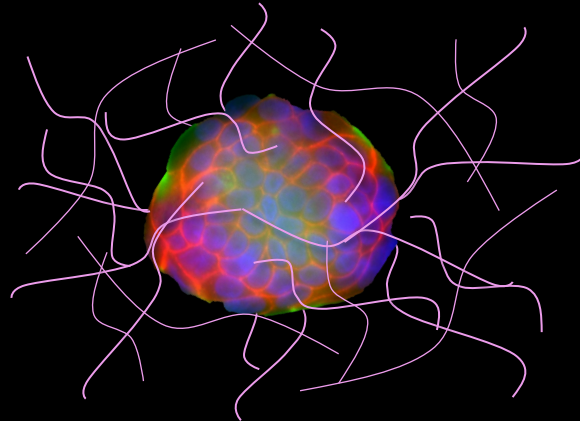
There is a 4th mode of cell migration, that is collective & amoeboid



3- Clinical applications: implement PDO to guide personalized medicine strategies

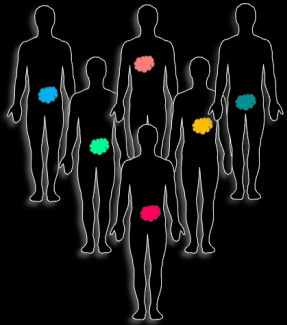
in Laminin-rich hydrogels

Matrigel derived from murin EHS tumors



Cancer patients require personalized medicine

Improve patient survival
& spare toxicities



Select from growing number
of therapeutic options



Match the right drug to the right patient



Genomic approaches only benefit 7-15% patients*

Functional precision medicine works for leukemias**

Are PDOs relevant tumor “avatars” for solid tumors?

*Le Tourneau et al., Lancet Oncol. 2015, Massard et al., Cancer Dis. 2017, Rodon et al., Nat medicine. 2019

**Kornauth et al. Cancer Dis. 2022 Malani et al., Cancer Dis. 2022

Missions

1

Implement PDO technology within patient clinical path & standardize the assays
Can we establish PDO for each patient? For the population?

2

Assess PDO clinical validity in this setting
How faithful are PDOs to the patient's tumor?

3

Determine the clinical utility of “functional” personalized medicine
How PDO can benefits to patient?

4

Leverage the pharmacotyping technology for all drugs
Expand current assays (chemograms) to immuno-oncology (immunograms)

anr



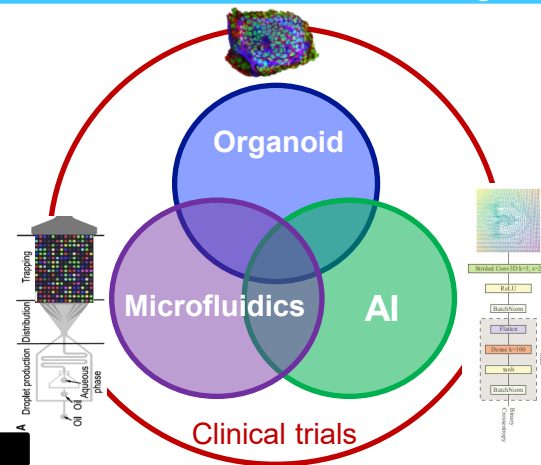
MINISTÈRE
DES SOLIDARITÉS
ET DE LA SANTÉ

PI: F Jaulin
Funded 2022-27

Consortium



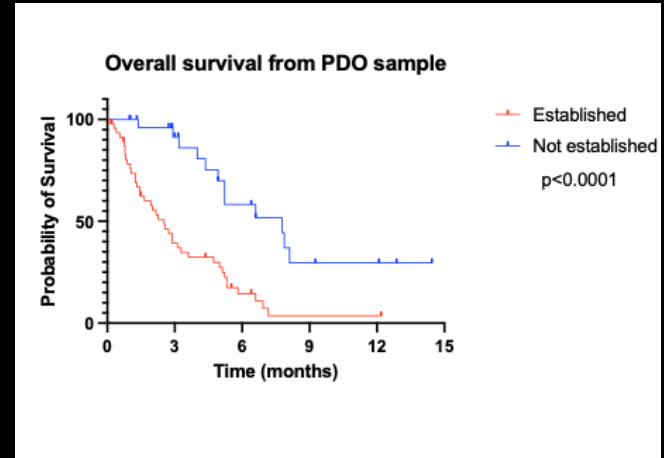
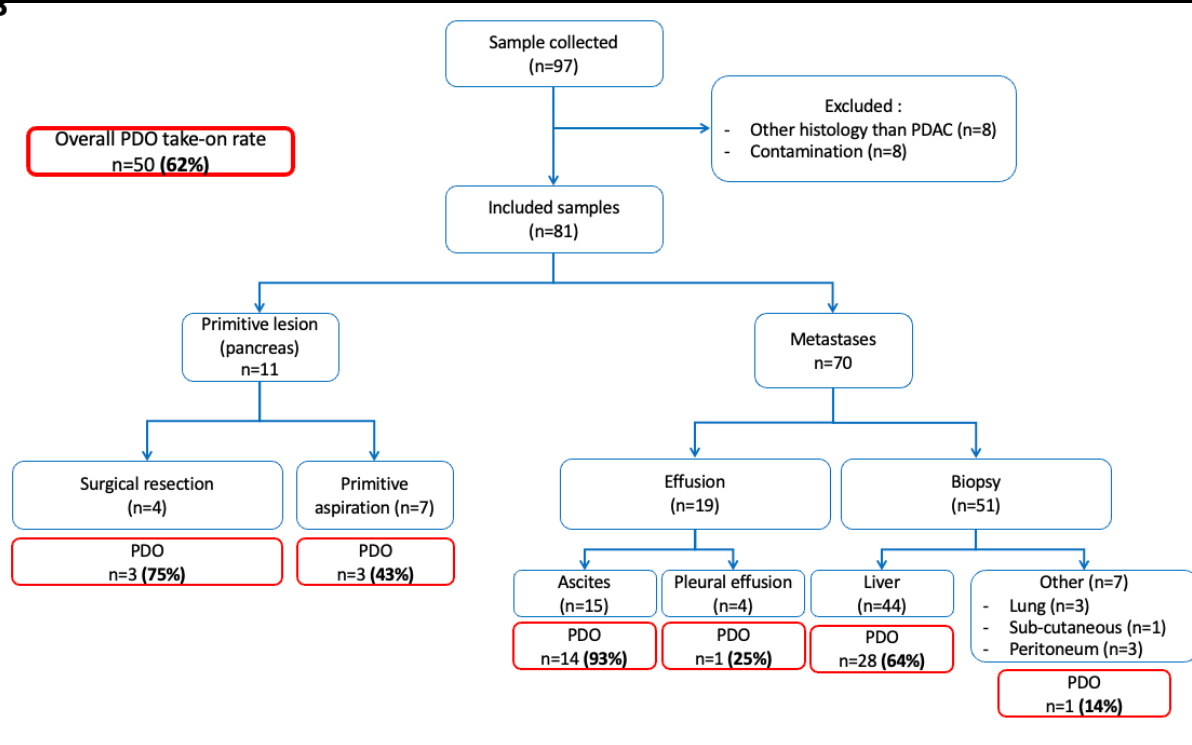
Unique combination of Expertise & ambitious clinical setting



Implement PDO technology within patient clinical path

1

We establish PDOs for all patients with different success rates

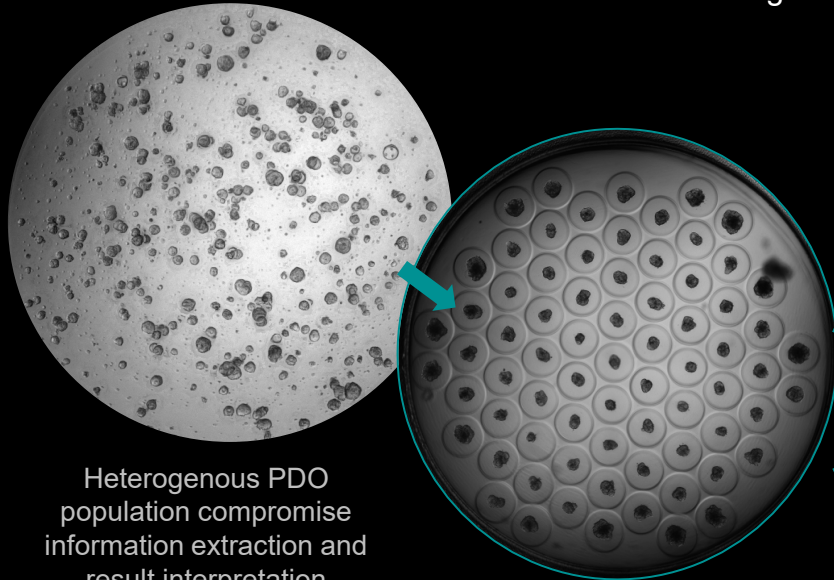


Standardize the assays & solve technological bottlenecks

1

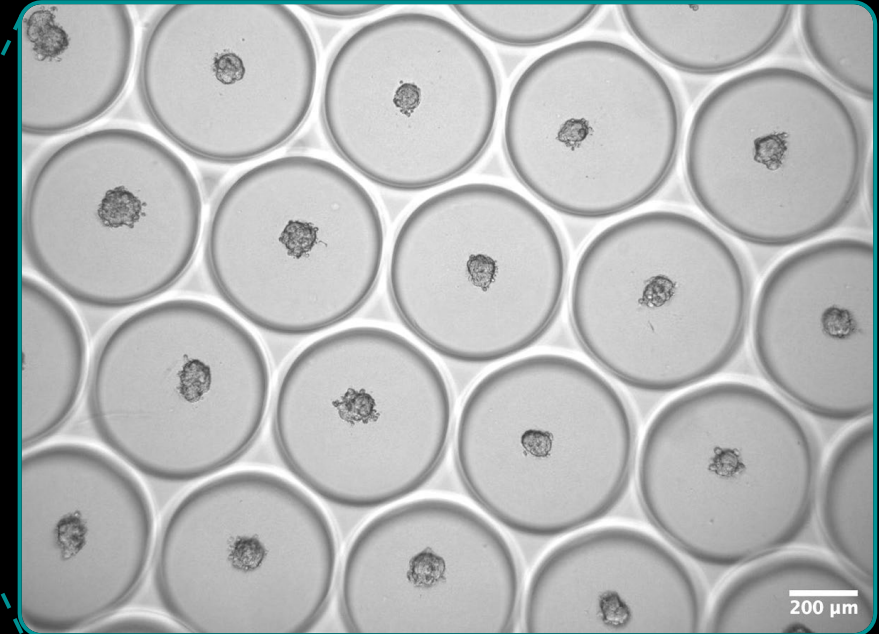
Time & space resolved microscopy

- To standardize, automatize for scaling-up screening capacities
- To resolve intratumoral heterogeneity
- To leverage artificial intelligence for upgraded read-outs (dynamic)



Heterogenous PDO
population compromise
information extraction and
result interpretation

Micro-engineering
(Microfluidics & microwells)

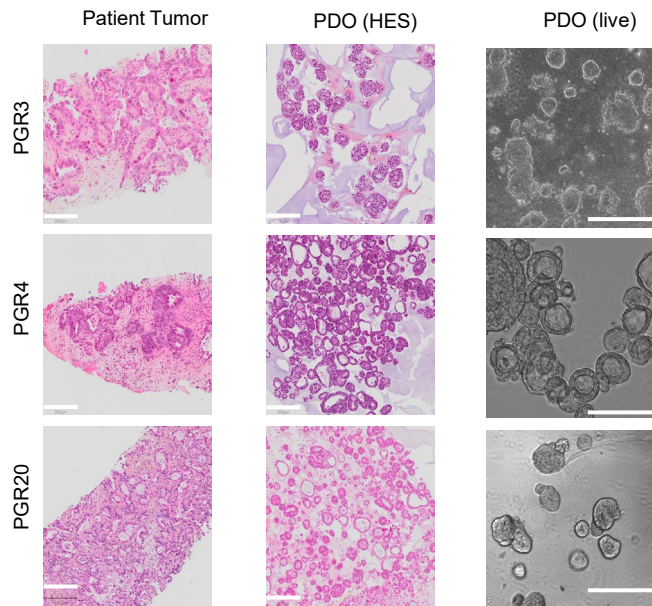


PDO from CRC patient in microwells over 4days (Jaulin lab, Gontran/Cartry/Bedja)

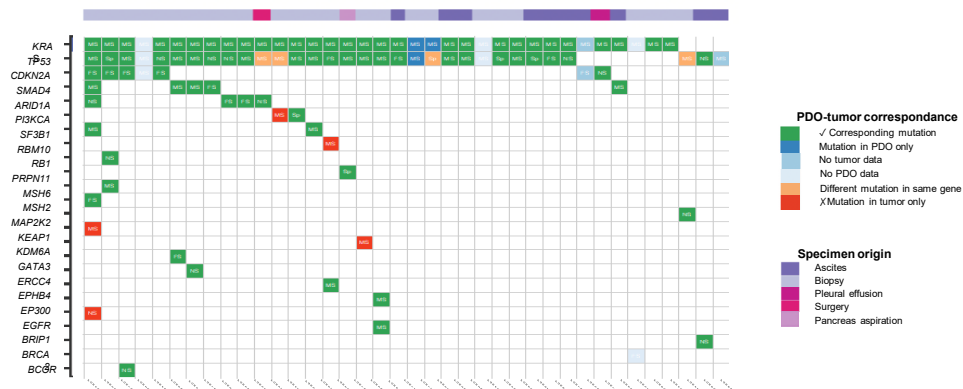
Biological concordance between PDOs & patient tumors

2

Histological concordance (cell autonomous multicellular organisation)



Genomic concordance (93%) (Early passages, MSS stable)



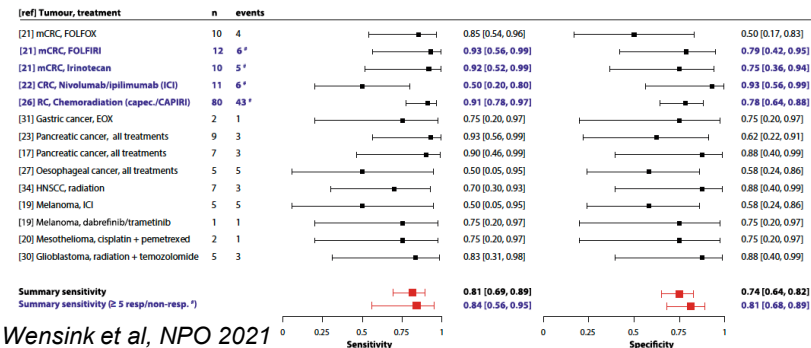
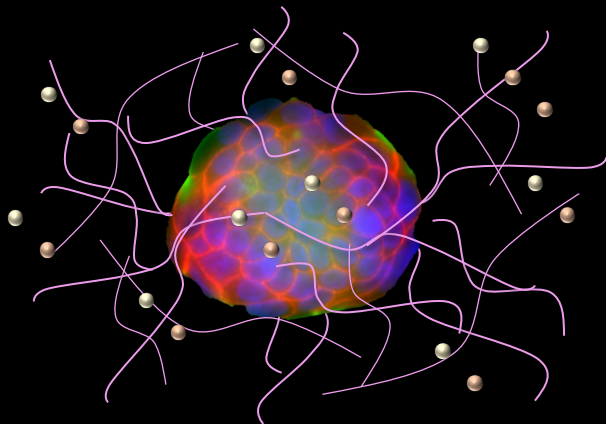
Exome sequencing from 40 tumors and associated organoids from pancreatic cancer patients

Clinical concordance

2

	PDAC (34 patients)	CRC (8 patients)
Sensitivity	83.3%	75%
Specificity	92.9%	75%
Positive pred. Value	71.4%	75%
Negative pred. Value	91.3%	75%

=> “Simple” PDO/matrigel culture predicts patient drug response to chemo & targeted therapies in clinical setting



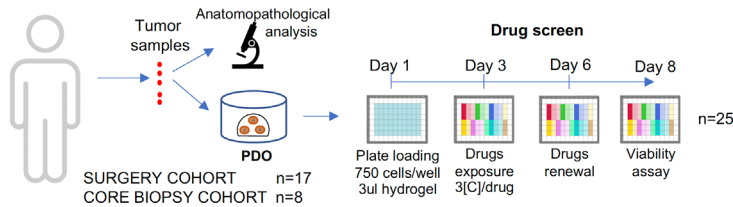
« Culture media composition influences patient-derived organoid ability to predict therapeutic responses in gastrointestinal cancers » *Hogenson et al, JCI insight 2022*

ORGANOTREAT Clinical trial

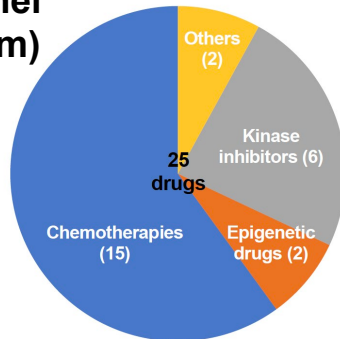
3

Interventional clinical trial aiming to orient patient treatment

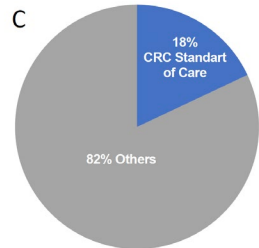
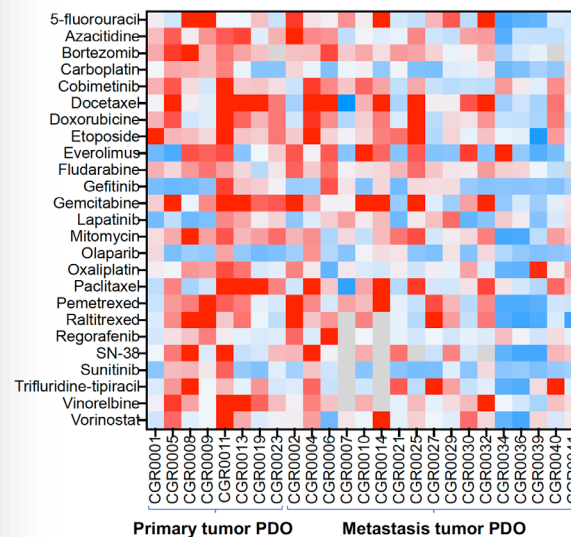
Chemogram



25-drug panel (Chemogram)



Drug response landscape in 25 patient pilot study



Could offer unexpected therapeutic options to patients

ORGANOTREAT Clinical trial

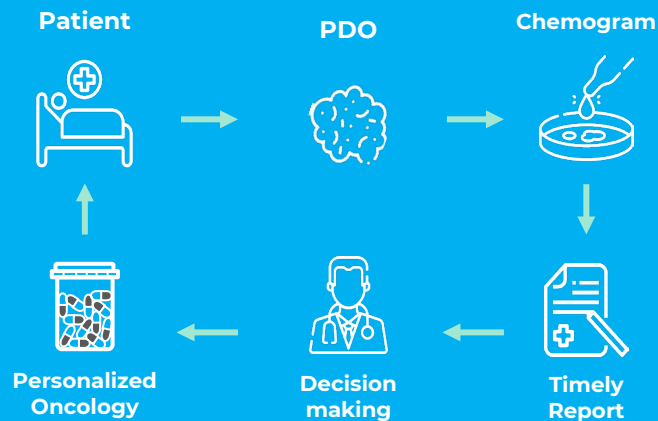
3

Prospective multicenter study evaluating the feasibility and efficacy of tumor **organoid-based precision medicine** in patients with advanced refractory **solid tumor**



	Phase :	Organ :	Patients
ORGANOTREAT-01	I/II	CRC	60 (GMI)
ORGANOTREAT-02R	III	PDAC	314
ORGANOTREAT-02R	III	CRC	582
ORGANOTREAT-02	II	Other	...

PI/Clinic: Pr Michel Ducreux, Alice Boileve

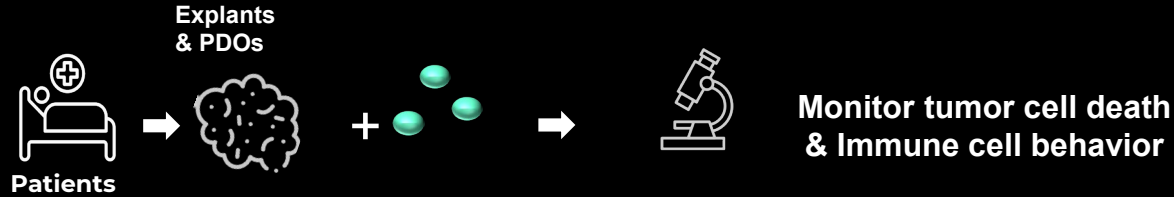


Interventional trial

Jerome Cartry and Sabrina Bedja

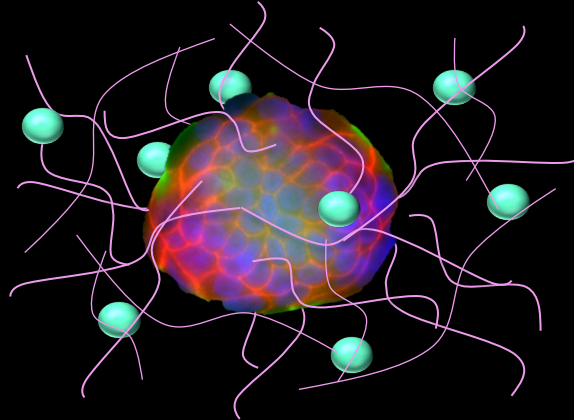
ALL solid tumors, >1000 patients

From chemograms to immunograms



1/ Engineered Car-T cells (Her-2/CD19)

2/ Autologous Immune cells (+/- ICB)



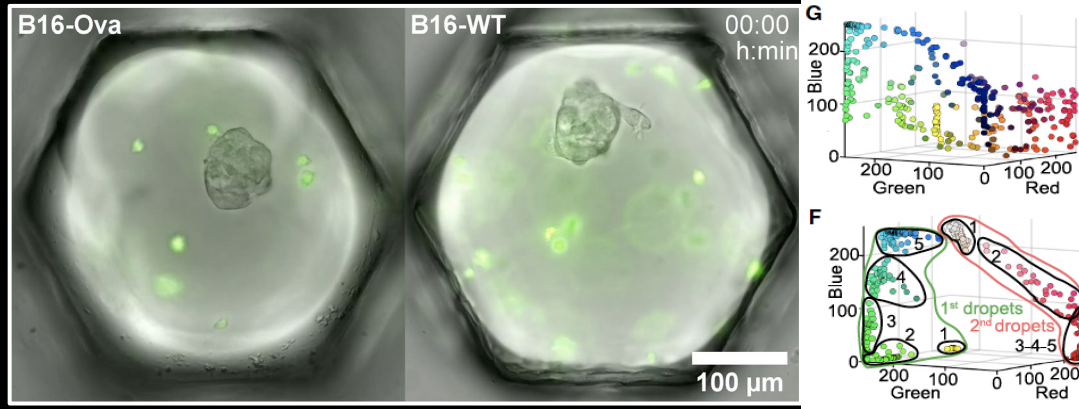
IMMUNOGRAM

Capture tumor immunogenicity
& predict response to Immune
Checkpoint Blockers (ICB)

Leverage the pharmacotyping technology for all drugs

4

From chemograms to immunograms



Ronteix et al. 2022

Charles Baroud, PhD
Group leader, Professor at Polytechnique
Physic-based approaches to droplet microfluidic



Philippe Bousso, PhD
Dir. Immunology department
Dynamic of immune response



Depend on:

« All Models are wrong
but some are useful »

- The question you ask
- The angle: holistic (relevance) or reductionist (mechanism, causality)
- The mission (cognitive, clinical application)
- Practical considerations (scalability, cost...)

We chose the patient-derived organoids as *ex vivo* tumor models (and happy about it!)

BUT, missing a lot of the tumor microenvironment

Adding specific cellular component will only **partially recover tissue complexity**

Restrict our understanding of disease, but will enable progresses



J. Cartry
Project leader

Patients!
(My) Dream Team !

U1279
Comité pathologies digestives
Département d'anatomopathologie
Radiologie interventionnelle
Pharmacie
PFEP, PFIC, PETRA
SEAT (Karelia Lipson/Negaar Goudarzi)

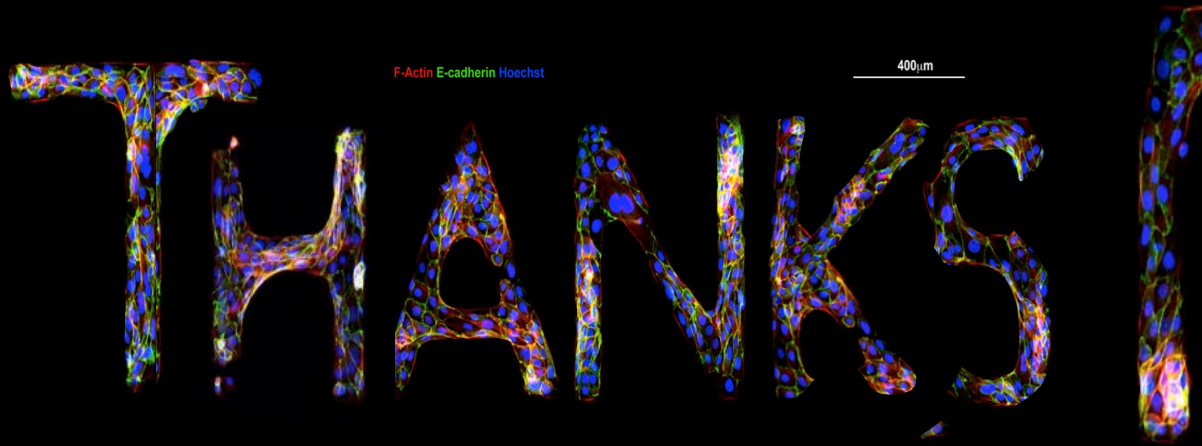
ACKNOWLEDGEMENTS

Collaborateurs

Matthieu Piel, IPGG, Paris
Raphael Voituriez, Sorbonne, Paris
Mathieu Coppey, Curie, Paris
Christophe Desterke, Paul Brousse
Vito Conte, UTE, Endhoven, Netherlands
Valeria Barresi, Italy
Julie Pannequin, IGF, Montpellier

Financements



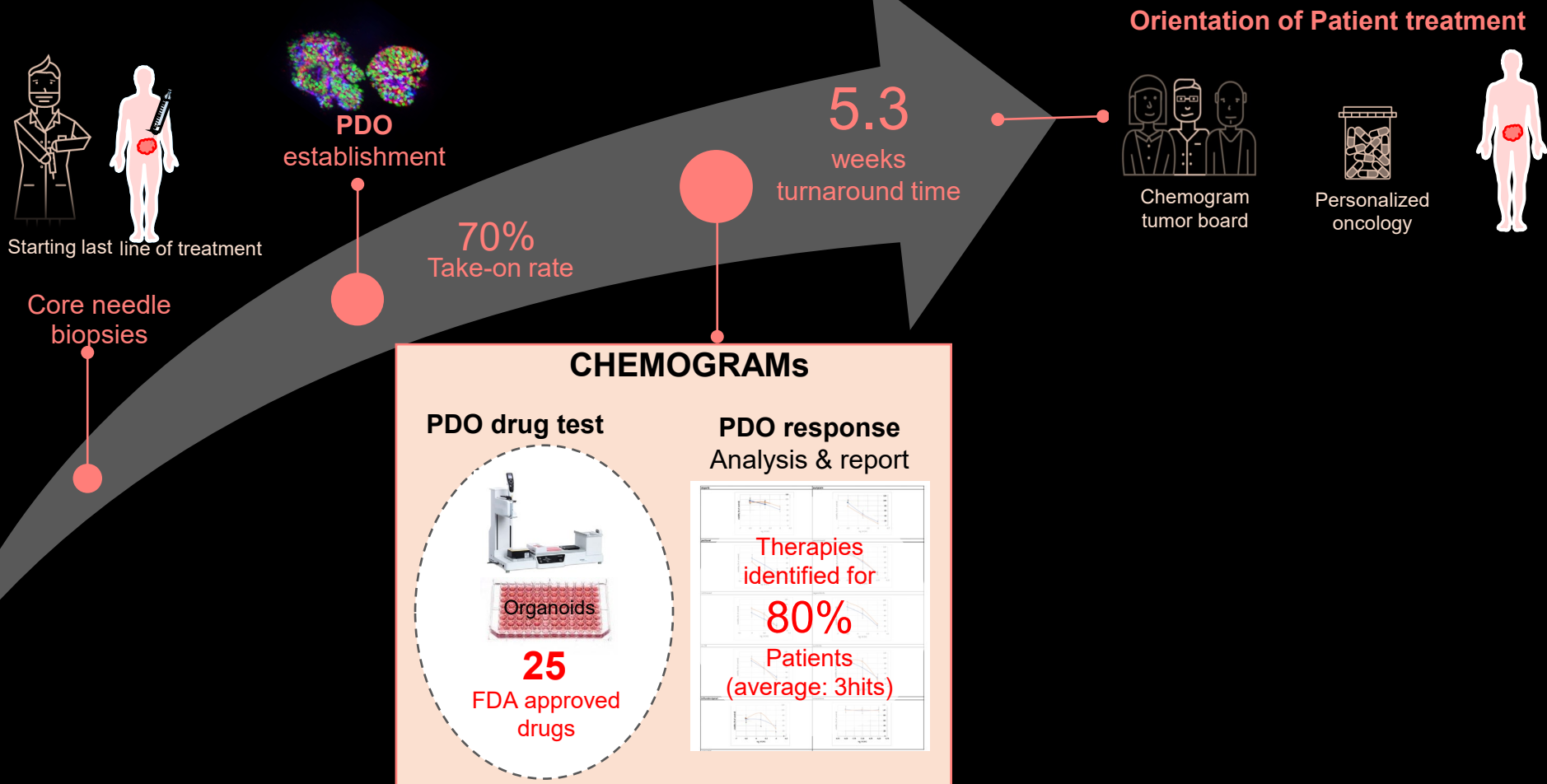


Jobs opportunities for Organoids fans!

- Postdoctoral position / RHU Organomic
- Senior Organoid Scientist / Orakl-oncology
- Head of Lab / Orakl-oncology

Send applications to fanny.Jaulin@gustaveroussy.fr

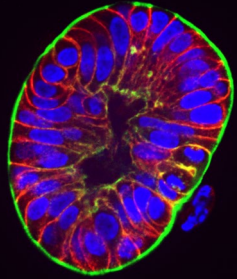
Functional personalized medicine



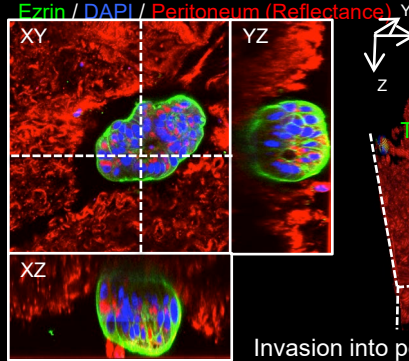


Collective migration without traction/adhesion???

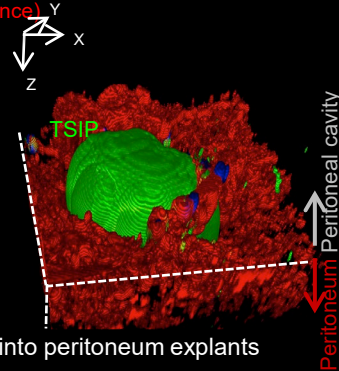
TSIP (Tumor Spheres with Inverted Polarity)



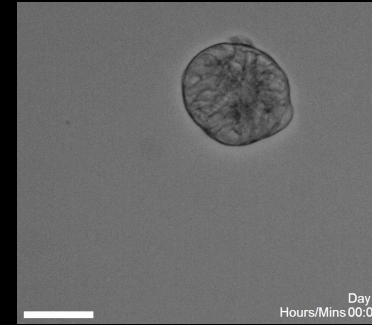
ITGB1/ Actin/ DAPI



Invasion into peritoneum explants



No protrusion? No leader?

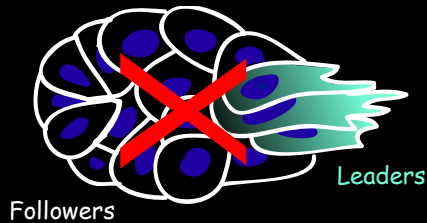


Migration into Collagen-I gels

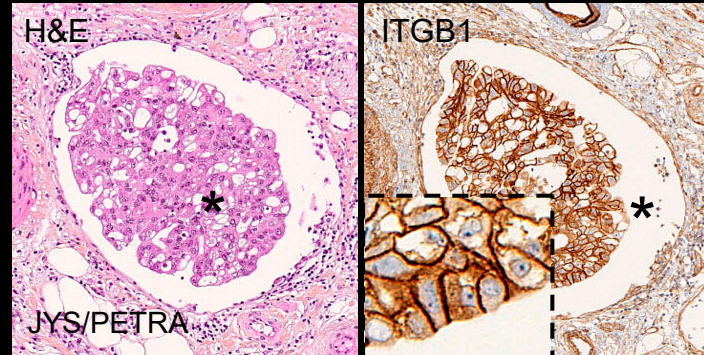
Collective

“Traction”

Adhesion/front



Mismatch between tumor cell adhesion repertoire & local ECM in the dissemination route



CRC tumour embolies into lymphatic vessels

Cell-autonomous

Non Cell-autonomous

Uncovering collective amoeboid migration



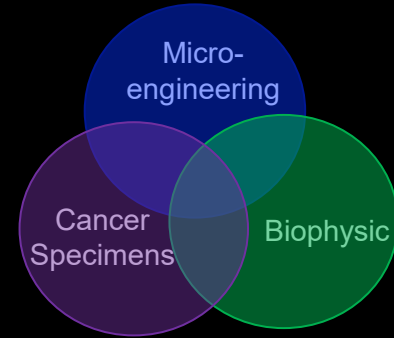
Matthieu Piel
IPGG/curie



Raphael Voituriez
Sorbonne University

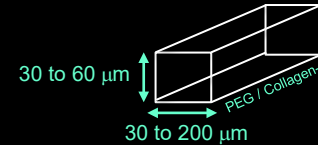


Mathieu Coppey
Curie Institute



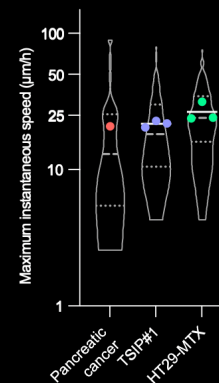
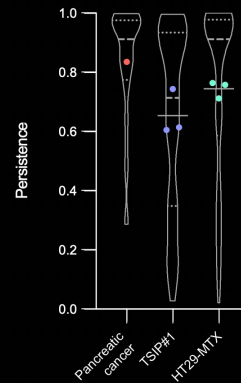
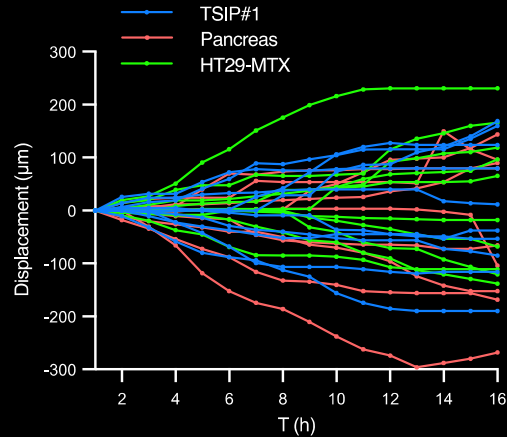
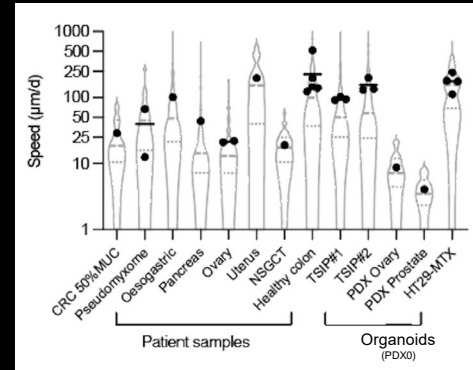
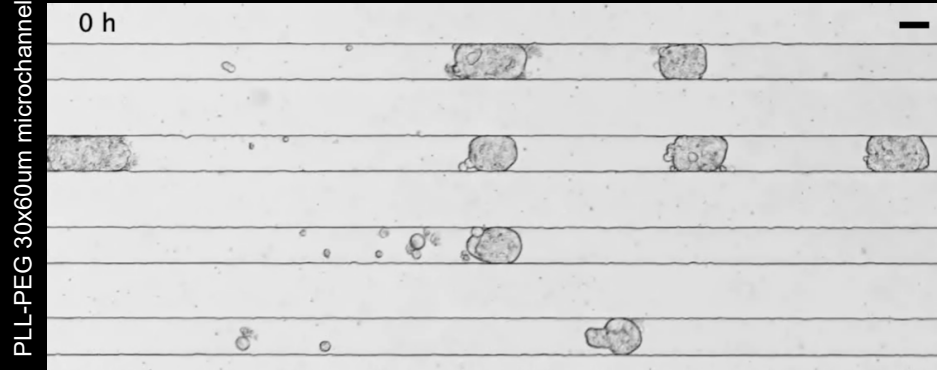
Microfluidics

Geometry / adhesion
(no gradient)



... from physicians to physicists!

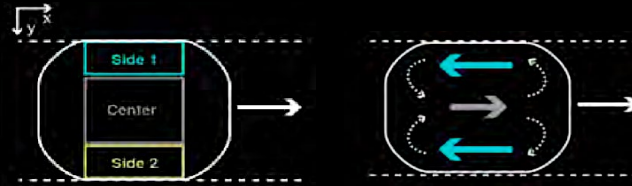
Most cancer cell clusters migrate into non-adhesive microchannels



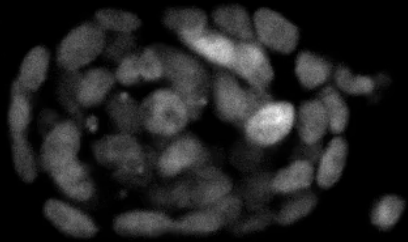
- No contribution of focal adhesion & traction forces
- Integrins participate in migration by exerting friction forces (In line with Berget et al. NCB 2015)

Cluster migration does not involve persistent retrograde flows of cells

How are propulsive forces generated?

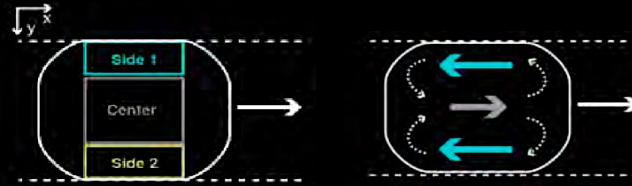


No neighbors exchange, clusters move as solids



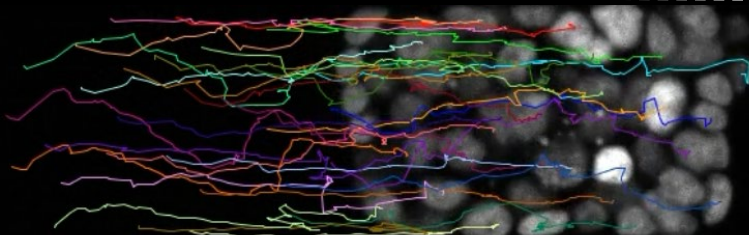
Cluster migration does not involve persistent retrograde flows of cells

How are propulsive forces generated?



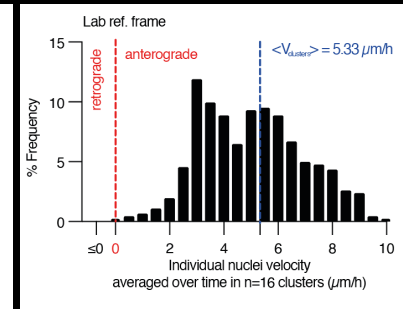
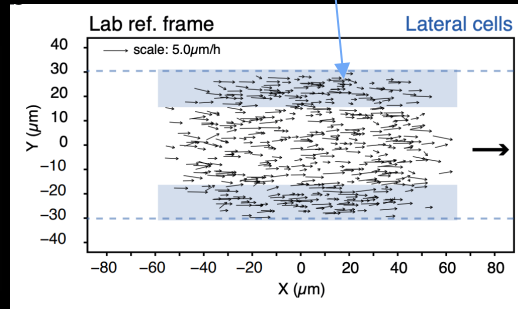
=> No cell Treadmilling !!

No neighbors exchange, clusters move as solids



HT-29 Cherry-H2B30x60 μm PLL-PEG

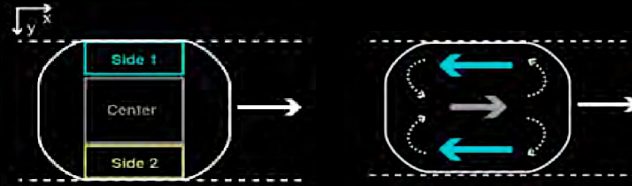
Anterograde flows



Average over time & clusters (n=16)

Cluster migration does not involve persistent retrograde flows

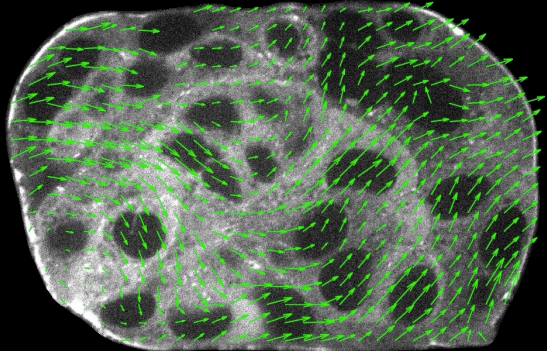
How are propulsive forces generated?



=> No persistent acto-myosin retrograde

0 min

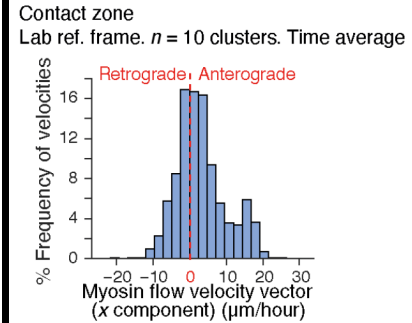
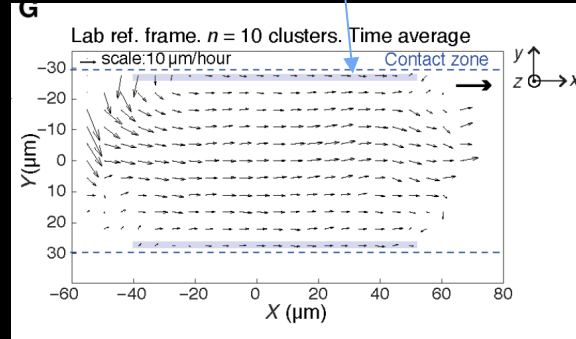
Particle Image Velocimetry



HT-29 mTurquoise MLC
30x60 μm PLL-PEG

20 μm

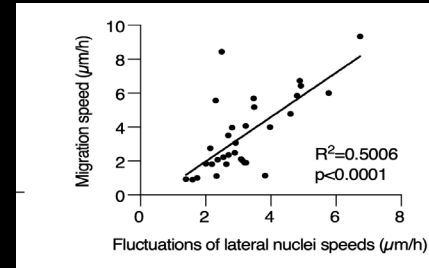
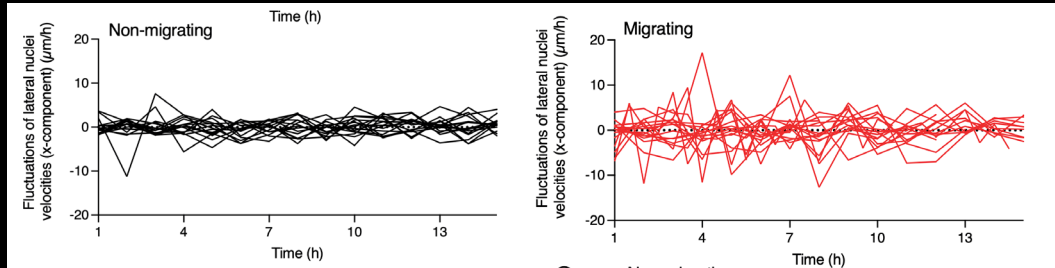
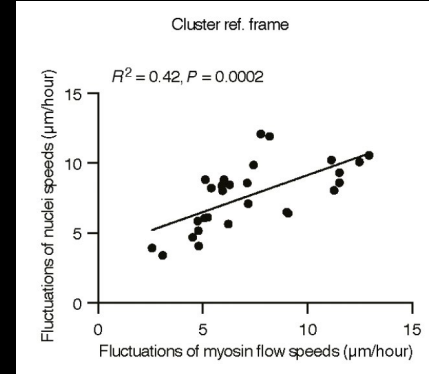
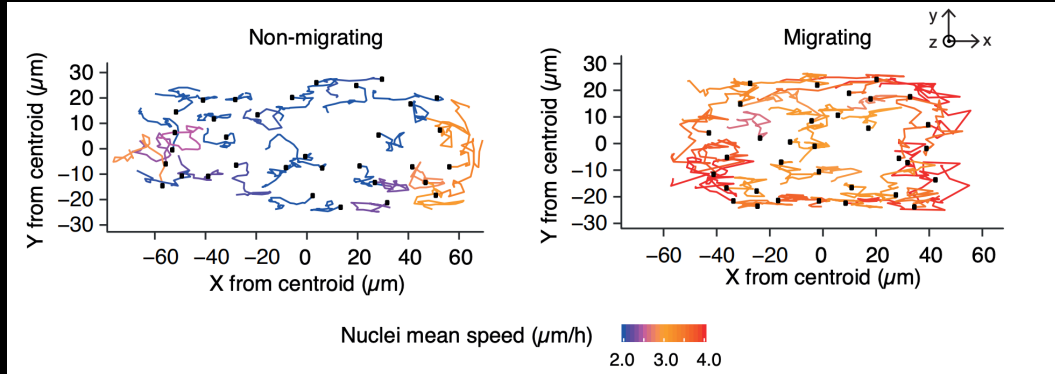
Anterograde flows



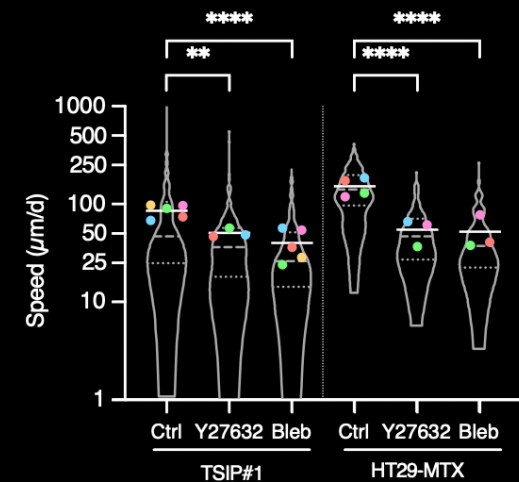
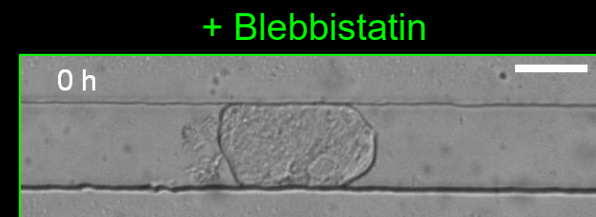
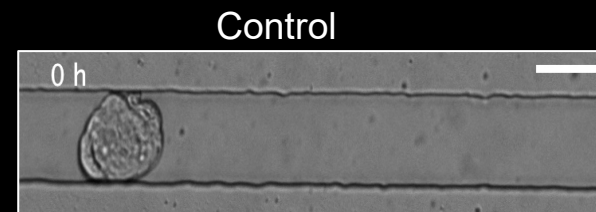
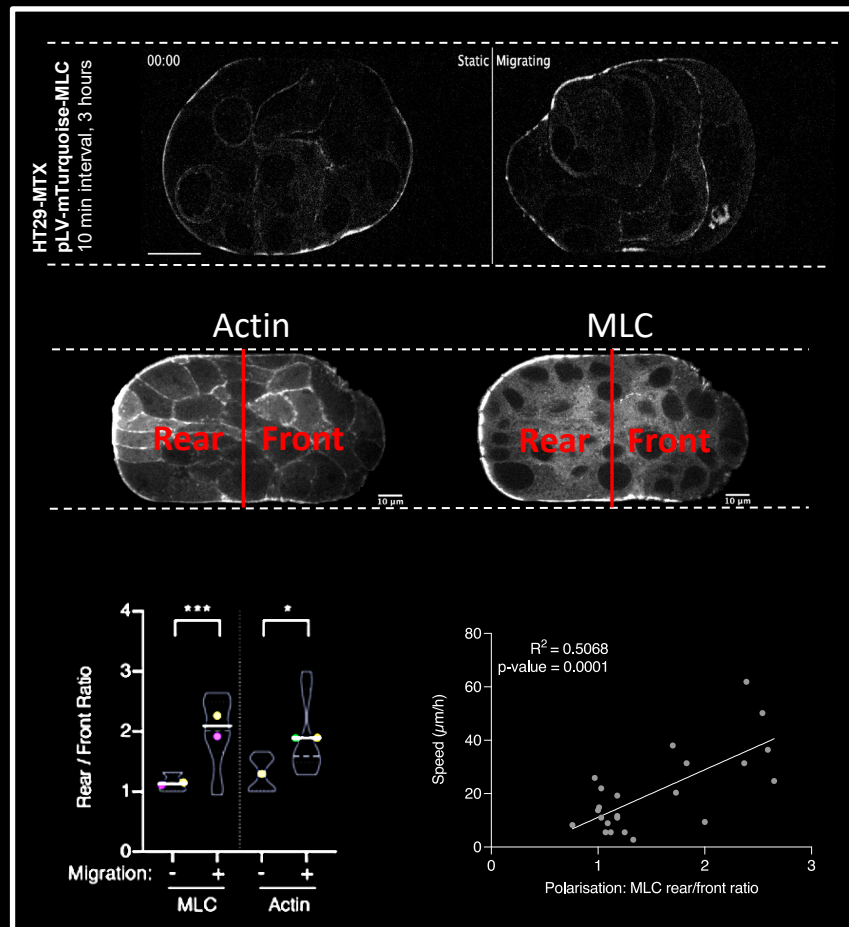
Average over clusters ($n=10$) & time (hours)

Fluctuation of myosin flow associated to cell « Jiggling »

Fluctuation of nuclei displacements



The Front/back polarization of the cortical acto-myosin cortex drives cluster migration



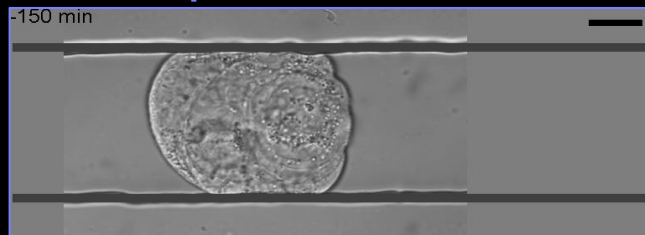
The Front/back polarization of the cortical acto-myosin cortex drives cluster migration

Optogenetic activation of Rho-A controls directionality

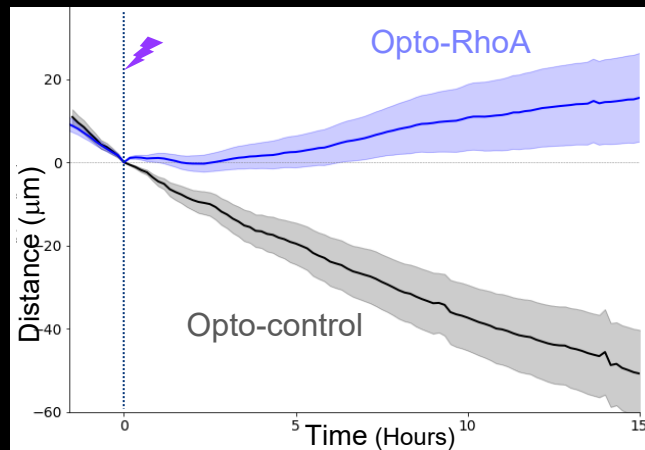
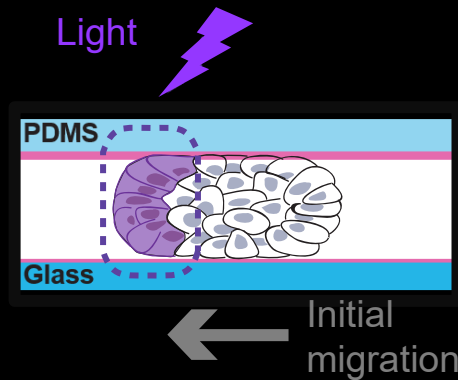
Opto-control



Opto-RhoA



HT29
CYBN-eGFP-CaaX
ARHGEF11CRY2-mCherryN1



Jean De Seze (X)
(Coppey lab)

There is a 4th mode of cell migration, not powered by persistent retrograde flows

